

CHAPTER 3

Construction of Vicinal Tertiary and All-Carbon Quaternary Stereocenters via Ir-Catalyzed Regio-, Diastereo-, and Enantioselective Allylic Alkylation and Applications in Sequential Pd-Catalysis¹

3.1 INTRODUCTION

3.1.1 State of the art in the asymmetric construction of vicinal quaternary and tertiary carbon centers

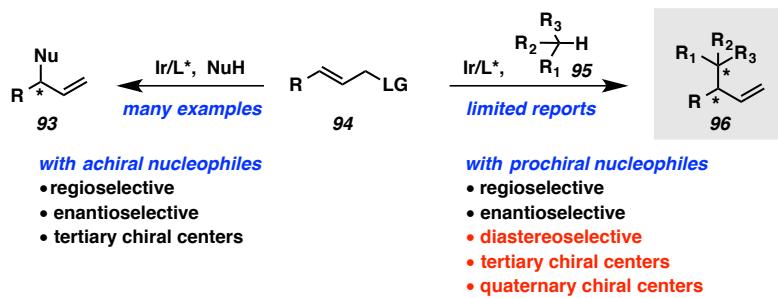
The asymmetric construction of sterically-encumbered, vicinal stereogenic centers is of great interest to synthetic chemists due to the prevalence of such structural arrangements in natural products and bioactive compounds.⁶⁹ The limited number of methods that provide selective access to vicinal tertiary and all-carbon quaternary stereocenters highlights the challenging nature of this task. Enantioselective approaches for accessing this structural dyad have generally relied on asymmetric Michael additions⁷⁰ and Claisen rearrangements.⁷¹ Among the methods available for forging this motif, only a relatively small number have been reported to do so by employing transition metals in a

¹ This work was performed in collaboration with Wen-Bo Lui, postdoctoral researcher in the Stoltz group, and Scott Virgil, manager of the Caltech Center for Catalysis and Chemical Synthesis. This work has been published. See: Liu, W. -B.; Reeves, C. M.; Virgil, S. C.; Stoltz, B. M. *J. Am. Chem. Soc.* **2013**, *135*, 10626.

catalytic, asymmetric fashion.^{72,73} Thus, further investigations into the development of metal-catalyzed methods to directly and selectively generate such stereochemical arrays should prove valuable.

Allylic alkylation chemistry represents a successful strategy for the assembly of highly congested chemical architectures by C–C bond formation⁷⁴ and, within this domain, Ir-catalyzed processes are among the most selective and highest yielding.^{75,76} Initial reports from the Helmchen⁷⁷ and Hartwig⁷⁸ groups demonstrated the utility of Ir-catalyzed allylic substitutions for the synthesis of enantioenriched 3,3-disubstituted (branched) allyl compounds.⁷⁵ As this research area has developed, Ir/phosphoramidite catalysts⁷⁹ have emerged as privileged scaffolds for the regio- and enantioselective allylic alkylation of achiral nucleophiles, such as malonate derivatives and ketone enolates (Figure 3.1.1.1, **94** → **93**).⁸⁰ However, methods for Ir-catalyzed intermolecular allylic alkylation that employ prochiral nucleophiles and display high (1) regio-, (2) diastereo-, and (3) enantioselectively remain elusive (Figure 3.1.1.1, **94** → **96**).⁸¹ Prior to our investigation, only two reports,⁸² from the laboratories of Takemoto and Hartwig, detailed successful examples in attaining all three of these goals; however, in these accounts, the nucleophiles investigated were limited to amino acid derivatives and azlactones.⁸³

Figure 3.1.1.1. Ir-catalyzed allylic substitution



In this chapter, we detail the development of a highly regio-, diastereo-, and enantioselective Ir-catalyzed α -allylic alkylation of cyclic β -ketoesters that forges vicinal tertiary and all-carbon quaternary centers in one step and in excellent yields. Moreover, we describe the deployment of our novel 2-(trimethylsilyl)ethyl β -ketoester (TMSE β -ketoester) (see Chapter 2), which functions as an oxycarbonyl-protected enolate, enabling sequential catalyst-controlled α -allylic alkylations and, in turn, the ability to select the diastereomer produced within the nascent stereochemical dyad.

3.2 REACTION DEVELOPMENT AND OPTIMIZATION

3.2.1 Discovery and optimization of iridium catalyzed regio-, diastereo- and enantioselective allylic alkylation of cyclic ketones

Our preliminary studies focused on probing the effects of different ligands, bases, additives, and solvents on the efficiency and selectivity of the reaction. Cyclic β -ketoester **97**, cinnamyl carbonate **98**, and $[\text{Ir}(\text{cod})\text{Cl}]_2$ /phosphoramidite complexes⁸⁴ were chosen as standard reaction components at the outset of our investigations.⁸⁵ Selected

results of these experiments are summarized in Table 3.2.1.1. Our investigations commenced with commonly used phosphoramidite ligand **L10**⁸⁶ and we were pleased to find that the proposed reaction proceeded smoothly under the conditions described (Table 3.2.1.1), delivering α -quaternary β -ketoester **100a** in >95% conversion and in 96% ee. Unfortunately, no diastereoselectivity was observed in this case (Table 3.2.1.1, entry 1). Use of ligand **L11**, a diastereoisomer of **L10**, again produced a high-yielding reaction, but in significantly diminished ee (32%) and modest 1:2 dr (entry 2). Inspired by the You group's use of Ir-*N*-arylphosphoramidite complexes (derived from $[\text{Ir}(\text{cod})\text{Cl}]_2$ and **L12**)⁸⁷ to effect the diastereo- and enantioselective intramolecular allylation of indoles and pyrroles,⁸⁸ we envisioned that analogous Ir complexes may prove valuable for the generation of all-carbon quaternary stereocenters. We were delighted to discover that the use of *N*-aryl-phosphoramidite ligand **L12** furnished the desired product in 98% ee, >20:1 dr, and 95:5 branched to linear ratio (entry 3).

Figure 3.2.1.1. Selected phosphoramidite and PHOX ligands

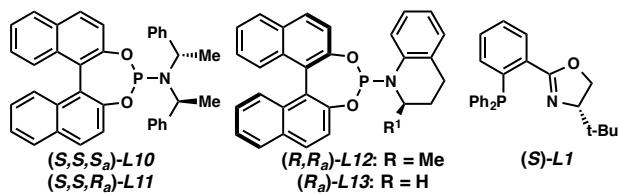


Table 3.2.1.1. Optimization of reaction parameters.^a

entry	<i>L</i>	base or additive (equiv)	100a:101a ^b	dr of 100a ^b	ee of 100a (%) ^c
1	<i>L10</i>	NaH (2)	>95:5	1:1	96 (99) ^d
2	<i>L11</i>	NaH (2)	>95:5	1:2	32 (3) ^e
3	<i>L12</i>	NaH (2)	95:5	>20:1	98
4	<i>L12</i>	-	80:20	11:1	96
5	<i>L12</i>	Et ₃ N (2)	77:23	11:1	97
6	<i>L12</i>	Cs ₂ CO ₃ (2)	63:37	6:1	93
7	<i>L12</i>	K ₃ PO ₄ (2)	63:37	4:1	90
8	<i>L12</i>	LiOt-Bu (2)	95:5	>20:1	99
9	<i>L12</i>	LiCl (1)	88:12	14:1	98
10	<i>L12</i>	LiBr (1)	95:5	>20:1	>99
11	<i>L13</i>	LiBr (1)	80:20	12:1	96
12 ^f	<i>L1</i>	LiBr (1)	12:88	-	-
13 ^g	<i>L12</i>	LiBr (1)	95:5	>20:1	99

^a Reactions performed with 0.1 mmol of **98a**, 0.2 mmol of **97a** at 0.1 M in THF at 20 °C and allowed to proceed to complete consumption of **98a**. ^b Determined by ¹H NMR and UHPLC-MS analysis of the crude mixture. ^c Determined by chiral HPLC analysis of the major diastereomer. ^d (ee) of the alternate diastereomer. ^e (ee) of the major diastereomer. ^f Measured after 60 h at 60% conversion. ^g 1 mol % [Ir(cod)Cl]₂ and 2 mol % **L12** were used.

Extensive exploration of various bases, including organic and inorganic bases, revealed that the use of LiOt-Bu afforded the desired product in comparable selectivities as NaH (entries 4–8). Previous reports demonstrating the marked effect of LiCl on the regioselectivity^{79b,80j,81b} in Ir-catalyzed allylic alkylations prompted us to investigate this and related additives. As a result of these efforts (entries 9–10), the combination of LiBr

and THF, at 25 °C, was found to provide **100a** in >20:1 dr, 95:5 branched:linear ratio, and with >99% ee (entry 10).

3.2.2 Further development of the reaction conditions

Under these superior conditions, several more ligands were examined. Use of ligand **L13**^{88b} afforded **100a** in 96% ee and 12:1 dr (entry 11). Employment of *N*-aryl-phosphoramidite scaffolds proved critical to maintaining high diastereoselectivity in the reaction. Phosphinooxazoline (PHOX) type ligands (e.g. **L1**), first used in Ir-catalyzed allylation by Helmchen,⁷⁷ were also examined, but we found these to be poorly suited for our reaction (entry 12). Finally, we found that the catalyst loading could be reduced to 1 mol % (entry 13) without loss of selectivity.

3.3 SURVEY OF REACTION SCOPE

3.3.1 Exploration of the reaction scope with respect to allyl electrophile

With optimized conditions in hand, the scope of substrates tolerated in the reaction was explored. We found that cinnamyl-derived carbonates bearing either electron-donating (–OMe) or electron-withdrawing (–Br, –CF₃) groups on the aromatic ring gave remarkably high dr, ee and yields (99% ee and 20:1 dr, Table 3.3.1.1, entries 1–4). The branched to linear ratio (i.e., **100:101**) tended to decrease as the electron deficiency of the aryl substituents increased (from 95:5 to 71:29, with 4-OMe-C₆H₄ to 4-CF₃-C₆H₄, respectively, entries 2–4). Heteroaryl substituents, such as 3-pyridyl, 2-thienyl and 2-furanyl, were also installed with uniformly excellent enantioselectivities and high diastereoselectivities (95–98% ee and 10:1–17:1 dr, entries 5–7). In addition to aromatic

substituents, methyl sorbyl carbonate was also well tolerated in the chemistry providing diene **100h**, although with a slight decrease in dr and ee (8:1 dr and 90% ee, entry 8).⁸⁹ Moreover, the reaction proceeded smoothly with ethyl β -ketoester **97i**, providing the α -allyl β -ketoester **100i** with excellent yield and selectivity (entry 9).

Figure 3.3.1.1. Substrate scope of Ir-catalyzed allylic alkylation of β -ketoesters^a

entry	product (100)	t (h)	100:101 ^b	dr of 100 ^b	yield (%) ^c	ee of 100 (%) ^d
1	100a: R ² = Ph	1	95:5	>20:1	98	>99
2	100b: R ² = p-MeO-C ₆ H ₄	1	95:5	20:1	99	>99
3	100c: R ² = p-Br-C ₆ H ₄	8	90:10	>20:1	90	99
4	100d: R ² = p-CF ₃ -C ₆ H ₄	12	71:29	>20:1	99	>99
5	100e: R ² = 3-pyridyl	12	50:50	11:1	98	98
6	100f: R ² = 2-thienyl	12	92:8	17:1	97	95
7	100g: R ² = 2-furanyl	12	95:5	10:1	90	95
8	100h: R ² = (E)-MeCH=CH	10	95:5	8:1	92	90
9	100i: R ¹ = Et	2	95:5	>20:1	98	>99

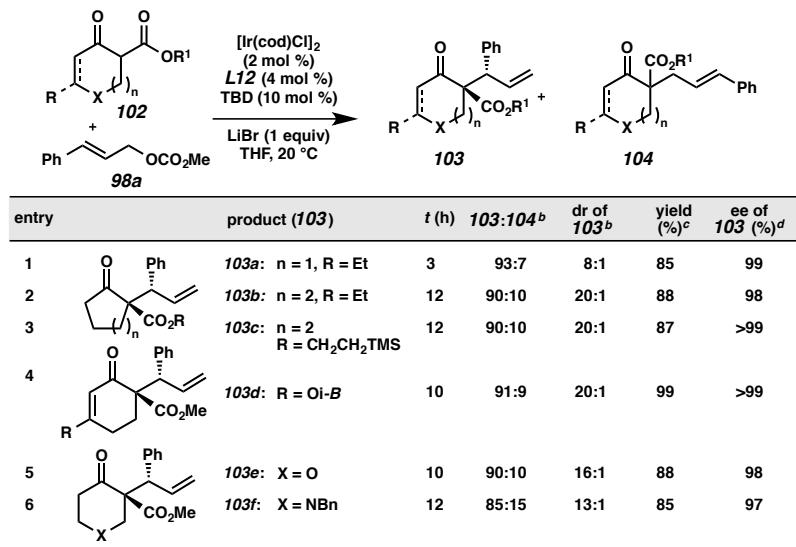
^a Reactions performed under the conditions of Table 1, entry 10. ^b Determined by ¹H NMR analysis of the crude mixture. ^c Isolated yield of **100** and **101**. ^d Determined by chiral HPLC or SFC analysis of the major diastereomer.

3.3.2 Exploration of the reaction scope with respect to ketoester nucleophile

Gratifyingly, aliphatic mono cyclic ketones also proved to be excellent participants in the reaction. Cyclopentanone and cyclohexanone based substrates delivered the products **103a** and **103b** in 98–99% ee and 8:1–20:1 dr, respectively (Figure

3.3.2.1, entries 1–2). Vinylogous ester, tetrahydropyran-4-one, and 4-piperidinone derivatives furnished the corresponding products (**103d–103f**) in high yields (85–99%), good diastereoselectivities (13:1–20:1), and enantioselectivities (97–99%, entries 4–6). The absolute stereochemistry of the product **103f** (>99% ee) was determined as (*R,R*) by single-crystal X-ray analysis.⁹⁰

Figure 3.3.2.1. Substrate scope of Ir-catalyzed allylic alkylation of β -ketoesters

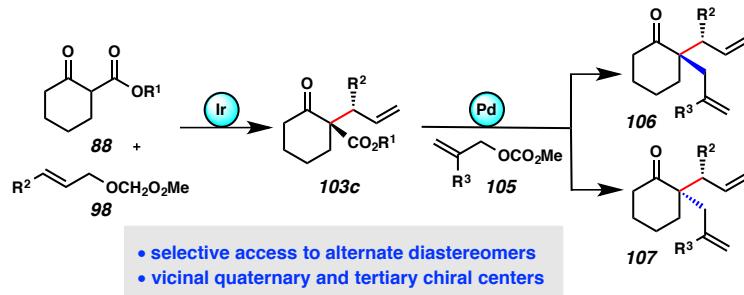


^a Reactions performed under the conditions of Table 1, entry 10. ^b Determined by ¹H NMR analysis of the crude mixture. ^c Isolated yield of **103** and **104**. ^d Determined by chiral HPLC or SFC analysis of the major diastereomer.

3.4 EMPLOYMENT OF TMSE- β -KETOESTERS TO ENABLE SEQUENTIAL CATALYSIS

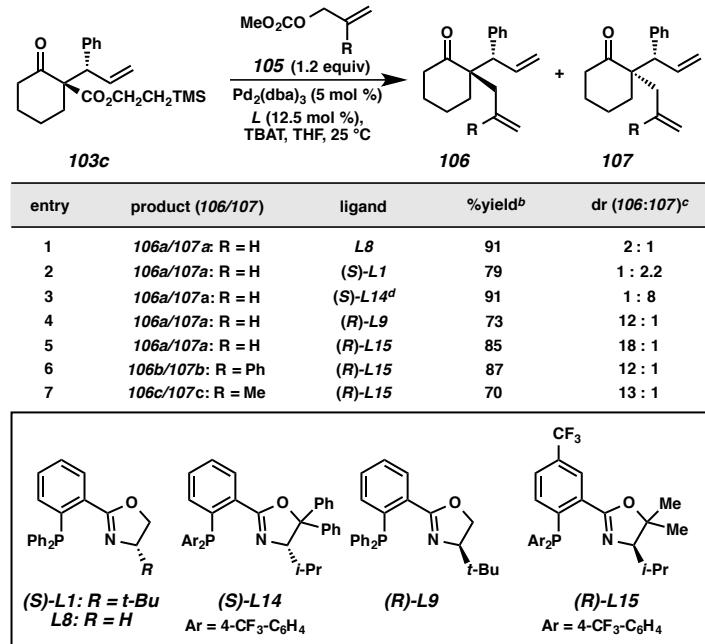
During the course of our investigations, we became intrigued by the possibility of developing a sequential allylic alkylation reaction, in which allylation of a dicarbonyl-stabilized enolate would be followed by palladium-catalyzed decarboxylative allylic alkylation and, thus, engender the ability to select among all four possible stereochemical outcomes.^{91,92} In order to realize such a consecutive allylic alkylation, we deployed our novel oxycarbonyl-protected enolate, TMSE β -ketoester **88**, which we hypothesized would successfully undergo Ir-catalyzed allylic alkylation and be poised to subsequently participate in Pd catalysis. Specifically, we envisioned that subsequent to Ir-catalyzed allylic alkylation TMSE β -ketoester **103c** could be triggered with fluoride, and the resulting prochiral enolate then intercepted and engaged in Pd-catalyzed allylic alkylation to deliver α -quaternary ketone **106** or **107** (Figure 3.4.1). In the case at hand, where β -ketoester **103c** contains a chiral branched R group at the a position, we anticipated that with careful choice of catalyst, we could potentially control the newly generated stereocenter independent of the absolute stereochemistry of the side chain.

Figure 3.4.1. Conceptualization of sequential catalysis



We were pleased to find that TMSE β -ketoester **88** is a highly competent substrate for Ir-catalyzed allylic alkylation and, under standard conditions, gave the desired product (**103c**) with excellent yield and selectivity (Figure 3.3.2.1, entry 3). Moreover, exposure of **103c** to catalytic $\text{Pd}_2(\text{dba})_3/\text{L8}$ (Figure 3.4.2) in the presence of allyl methylcarbonate and tetrabutylammonium difluorotriphenylsilicate (TBAT) generated the desired diallylated α -quaternary ketones **106** and **107** in good yield. The use of achiral PHOX ligand **L8** revealed that substrate **103c** displays inherent selectivity under Pd catalysis, furnishing **106a**⁹³ as the major diastereomer in a 2:1 ratio with **107a** (Figure 3.4.2, entry 1). Use of (*S*)-*t*-BuPHOX ligand (*S*)-**L1** resulted in modest reversal of the inherent diastereoselectivity to generate **107a** predominantly (entry 2). Furthermore, we were interested to find that use of ligand (*S*)-**L14**, possessing both an electronically modified phosphine and a smaller *i*-Pr substituent on the oxazoline ring in contrast to the more standard *t*-Bu, produced **107a** with improved diastereoselectivity (**106a:107a**, 1:8 dr) and 91% yield (entry 3). Alternatively, through judicious choice of ligand (e.g., (*R*)-**L15**), the inherent selectivity of the system could be enhanced to afford **106a:107a** with up to 18:1 dr favoring **106a** (entries 4–5). Cursory investigation revealed that 2-aryl and 2-alkyl substitutions at the allyl carbonate are well tolerated: allylic alkylation products **106b** and **106c** were obtained in good yields and with excellent diastereoselectivities.

Figure 3.4.2. Development of Pd-catalyzed diastereoselective decarboxylative allylic alkylation of TMSE- β -ketoesters^a



^a Reactions performed with 1.2 equiv of TBAT, and 1.2 equiv of allyl methylcarbonate at 0.03 M.^b

Isolated yield of **106** and **107**. ^c Determined by ¹H NMR analysis of the crude mixture and confirmed by GC analysis. ^d 10 mol % ligand was used.

3.5 CONCLUDING REMARKS

In summary, a highly regio-, diastereo-, and enantio-selective method for the synthesis of vicinal tertiary and all-carbon quaternary centers was realized through the use of an $[\text{Ir}(\text{cod})\text{Cl}]_2/\text{N-aryl-phosphoramidite}$ (**L12**) catalyst system. Varied substitutions were well tolerated on both the β -ketoester and allyl carbonate fragments. A sequential Ir/Pd-catalyzed dialkylation protocol was also established to deliver bis-allylated α -quaternary ketones with excellent stereoselectivity, while affording access to either product diastereomer with catalyst control. Further studies exploring the

mechanisms of these reactions and exploiting their applications in the total synthesis of complex natural products are underway in our laboratory.

3.6 EXPERIMENTAL SECTION

3.6.1 Materials and Methods

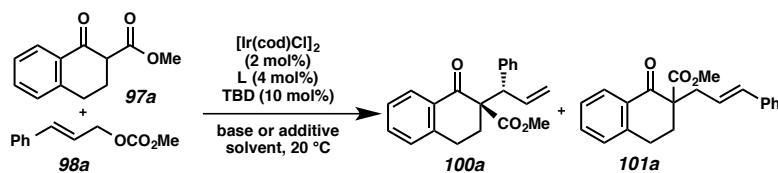
Unless otherwise stated, reactions were performed in flame-dried glassware under an argon or nitrogen atmosphere using dry, deoxygenated solvents. Solvents were dried by passage through an activated alumina column under argon.⁶¹ Reaction progress was monitored by thin-layer chromatography (TLC) or Agilent 1290 UHPLC-LCMS. TLC was performed using E. Merck silica gel 60 F254 precoated glass plates (0.25 mm) and visualized by UV fluorescence quenching, *p*-anisaldehyde, or KMnO₄ staining. Silicycle SiliaFlash® P60 Academic Silica gel (particle size 40–63 nm) was used for flash chromatography. ¹H NMR spectra were recorded on Varian Inova 500 MHz and 600 MHz spectrometers and are reported relative to residual CHCl₃ (δ 7.26 ppm) or C₆HD₅ (δ 7.16 ppm). ¹³C NMR spectra were recorded on a Varian Inova 500 MHz spectrometer (125 MHz) and are reported relative to CHCl₃ (δ 77.16 ppm) or C₆HD₅ (δ 128.06 ppm). ³¹P and ¹⁹F NMR spectra were recorded on a Varian Mercury 300 MHz (at 121 MHz and 282 MHz, respectively). ¹⁹F NMR spectra were reported relative to CFCl₃ (δ 0.0 ppm). ³¹P NMR spectra were reported relative to external H₃PO₄ (δ 0.0 ppm). Data for ¹H NMR are reported as follows: chemical shift (δ ppm) (multiplicity, coupling constant (Hz), integration). Multiplicities are reported as follows: s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, sept = septuplet, m = multiplet, br s = broad singlet, br d = broad doublet, app = apparent. Data for ¹³C NMR are reported in terms of chemical shifts

(δ ppm). IR spectra were obtained by use of a Perkin Elmer Spectrum BXII spectrometer using thin films deposited on NaCl plates and reported in frequency of absorption (cm^{-1}). Optical rotations were measured with a Jasco P-2000 polarimeter operating on the sodium D-line (589 nm), using a 100 mm path-length cell and are reported as: $[\alpha]_D^T$ (concentration in g/100 mL, solvent). Analytical HPLC was performed with an Agilent 1100 Series HPLC utilizing a Chiralpak (AD-H or AS) or Chiralcel (OD-H, OJ-H, or OB-H) columns (4.6 mm x 25 cm) obtained from Daicel Chemical Industries, Ltd. Analytical SFC was performed with a Mettler SFC supercritical CO_2 analytical chromatography system utilizing Chiralpak (AD-H, AS-H or IC) or Chiralcel (OD-H, OJ-H, or OB-H) columns (4.6 mm x 25 cm) obtained from Daicel Chemical Industries, Ltd. High resolution mass spectra (HRMS) were obtained from Agilent 6200 Series TOF with an Agilent G1978A Multimode source in electrospray ionization (ESI+), atmospheric pressure chemical ionization (APCI+), or mixed ionization mode (MM: ESI-APCI+).

Reagents were purchased from Sigma-Aldrich, Acros Organics, Strem, or Alfa Aesar and used as received unless otherwise stated. Ligands **L12–L13**,⁸⁷ ligands **L8–L9** and **L14–L15**,⁹⁴ allyl carbonates,⁹⁵ and β -ketoesters⁹⁶ were prepared by known methods.

3.6.2 Optimization of reaction parameters

Table 3.6.2.1. Optimization of reaction parameters



entry ^a	L	base/additive (equiv)	solvent	t (h)	conv (%) ^b	100a:101a ^b	dr of 100a ^b	ee of 100a (%) ^c
1	L10	NaH (2)	THF	24	>95	>95:5	1:1.4	96 (99) ^d
2	L11	NaH (2)	THF	60	>95	>95:5	1:1.9	32 (3) ^e
3	L12	NaH (2)	THF	12	>95	95:5	>20:1	98
4	L12	—	THF	8	>95	80:20	11:1	96
5	L12	DABCO (2)	THF	8	>95	72:28	5.0:1	96
6	L12	TBD (2)	THF	8	>95	74:26	11:1	95
7	L12	Et ₃ N (2)	THF	8	>95	77:23	11:1	97
8	L12	Cs ₂ CO ₃ (2)	THF	12	>95	63:37	6.3:1	93
9	L12	K ₃ PO ₄ (2)	THF	12	>95	63:37	4.1:1	90
10	L12	NaHMDS (2)	THF	12	>95	75:25	8.3:1	93
11	L12	LiHMDS (2)	THF	12	>95	86:14	13:1	96
12	L12	LiOt-Bu (2)	THF	1	>95	95:5	>20:1	99
13	L12	LiCl (1)	THF	1	>95	88:12	14:1	98
14	L12	LiBr (1)	THF	1	>95 (98)	95:5	>20:1	>99
15	L12	LiI (1)	THF	1	>95	72:28	>20:1	97
16	L10	LiBr (1)	THF	60	<5	nd	nd	nd
17	L11	LiBr (1)	THF	60	<5	nd	nd	nd
18	L13	LiBr (1)	THF	12	>95	80:20	12:1	96
19	L1	LiBr (1)	THF	60	60	12:88	nd	nd
20	L12	LiBr (1)	p-dioxane	1	>95	95:5	>20:1	>99
21	L12	LiBr (1)	Et ₂ O	3	>95	76:24	11:1	96
22	L12	LiBr (1)	CH ₂ Cl ₂	60	55	68:32	9.0:1	66
23	L12	LiBr (1)	toluene	16	>95	>95:5	>20:1	91
24 ^f	L12	LiBr (1)	THF	12	>95	95:5	>20:1	99
25 ^g	L12	LiBr (1)	THF	60	60	92:8	>20:1	94

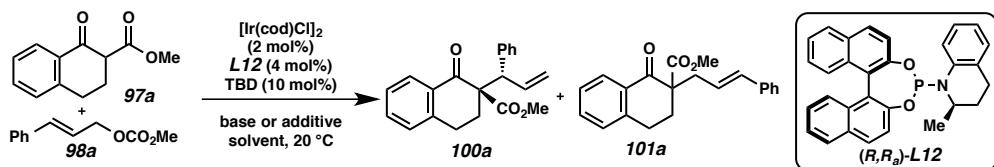
^a Reactions performed with 0.1 mmol of **98a**, 0.2 mmol of **97a** in 1 mL of solvent. ^b Determined by ¹H NMR or UHPLC-MS analysis of the crude reaction mixture. ^d (Ee) of the alternate diasteromer. ^e Measured on the minor isomer and the number in the parenthesis is ee of the major isomer. ^f 1 mol % of [Ir(cod)Cl]₂ and 2 mol % of **L12** were used. ^g 0.5 mol % of [Ir(cod)Cl]₂ and 1 mol % of **L12** were used.

General Procedure for Optimization Reaction (Table 3.6.2.1): All experiments were preformed in a nitrogen-filled glove box. [Ir(cod)Cl]₂ (1.4 mg, 0.002 mmol, 2 mol%), ligand (0.004 mmol, 4 mol%), and TBD (1.4 mg, 0.01 mmol, 10 mol%) were

added to a vial equipped with a magnetic stirring bar. The vial was then charged with solvent (0.5 mL) and stirred at 20 °C for 10 min, generating an orange solution. Cinnamyl carbonate **98a** (19.2 mg, 0.1 mmol, 1.0 equiv), β -ketoester **97a** (40.4 mg, 0.2 mmol, 2.0 equiv), base or additive (as indicated below) and another 0.5 mL of solvent were added. The vial was sealed and stirred at 20 °C until allylic carbonate **98a** was fully consumed, as indicated by TLC or UHPLC-MS analysis. The reaction mixture was filtered through a celite pad, rinsed with CH_2Cl_2 , and concentrated under reduced pressure. The ratios of constitutional isomers (branched product to linear product: **100a**:**101a**) and diastereomers (dr) were determined by ^1H NMR or UHPLC-MS.

3.6.3. General procedure for the Ir-catalyzed asymmetric allylic alkylation of β -ketoesters

Note: the absolute configuration was determined only for compound **100f** via X-ray analysis (vide infra, Appendix 4). The absolute configuration for all other products **100** has been inferred by analogy. Isolated yields are reported in Figures 3.3.1.1 and 3.3.2.1 (vida infra). For respective HPLC or SFC conditions, please refer to Table 3.6.8.1.



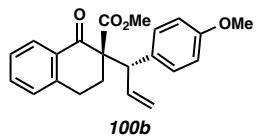
(R)-methyl 1-oxo-2-((S)-1-phenylallyl)-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100a). In a nitrogen-filled glove box, $[\text{Ir}(\text{cod})\text{Cl}]_2$ (2.7 mg, 0.004 mmol, 2 mol%), ligand **L12** (3.7 mg, 0.008 mmol, 4 mol%), and TBD (2.8 mg, 0.02 mmol, 10 mol%) were

added to a 2 dram scintillation vial equipped with a magnetic stirring bar. The vial was then charged with THF (1 mL) and stirred at 20 °C for 10 min, generating an orange solution. Cinnamyl carbonate (**98a**) (38.3 mg, 0.2 mmol, 1.0 equiv), LiBr (17.3 mg, 0.2 mmol, 1.0 equiv), β -ketoester **97a** (80.8 mg, 0.4 mmol, 2.0 equiv) and another 1 mL of THF were added. The vial was sealed and stirred at 20 °C until allylic carbonate **98a** was fully consumed, as indicated by TLC or UHPLC-MS analysis. THF was evaporated and the crude mixture was then dissolved in CH₂Cl₂, filtered through a celite pad, rinsed with CH₂Cl₂, and concentrated under reduced pressure. The regioselectivity (branched product to linear product: b:l = 95:5) and diastereoselectivity (dr >20:1) were determined by ¹H NMR or UHPLC-MS. The residue was purified by silica gel flash chromatography (gradient elution, 2→5% EtOAc in hexanes) to afford **100a** and **101a** (62.6 mg, 98% combined yield). Allylation product **100a** was isolated as a white solid by silica gel chromatography (gradient elution, 0→2% EtOAc in hexanes). >99% ee, $[\alpha]_D^{25} +26.3$ (c 1.11, CHCl₃); R_f = 0.3 (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 8.03 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.42–7.39 (m, 3H), 7.28–7.23 (m, 3H), 7.19–7.13 (m, 2H), 6.36 (dt, *J* = 16.8, 10.0 Hz, 1H), 5.21–5.12 (m, 2H), 4.46 (d, *J* = 10.0 Hz, 1H), 3.56 (s, 3H), 3.23 (ddd, *J* = 17.1, 12.1, 4.7 Hz, 1H), 2.88 (ddd, *J* = 17.6, 5.0, 3.0 Hz, 1H), 2.60 (ddd, *J* = 13.7, 4.7, 3.0 Hz, 1H), 2.10 (ddd, *J* = 13.6, 12.1, 5.0 Hz, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 193.2, 170.0, 143.1, 139.8, 136.6, 133.6, 132.5, 130.2, 128.8, 128.4, 128.2, 126.9, 126.7, 117.9, 62.7, 53.9, 52.6, 28.8, 26.4; IR (Neat Film, NaCl) 3066, 3028, 2948, 1731, 1685, 1636, 1599, 1491, 1453, 1433, 1358, 1298, 1283, 1238, 1214, 1169, 1108, 1080, 1032, 1001, 980, 926, 892, 808, 743 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd

for $C_{21}H_{21}O_3$ $[M+H]^+$: 321.1485, found 321.1489; HPLC conditions: 2% IPA, 0.6 mL/min, Chiralcel OD-H column, $\lambda = 254$ nm, t_R (min): major = 13.80, minor = 17.89.

3.6.4. Spectroscopic data for Ir-catalyzed allylic alkylation products

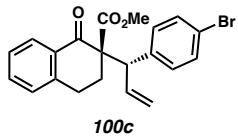
(R)-methyl 2-((S)-1-(4-methoxyphenyl)allyl)-1-oxo-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100b)



Ketoester **100b** was isolated by silica gel chromatography (gradient elution, 0→5% EtOAc in hexanes) as a white solid. >99% ee, $[\alpha]_D^{25} +38.5$ (c 0.93, $CHCl_3$); $R_f = 0.3$ (5% EtOAc in hexanes); 1H NMR (500 MHz, $CDCl_3$) δ 8.03 (dd, $J = 7.9, 1.4$ Hz, 1H), 7.41 (td, $J = 7.5, 1.5$ Hz, 1H), 7.37–7.31 (m, 2H), 7.31–7.21 (m, 1H), 7.21–7.12 (m, 1H), 6.86–6.75 (m, 2H), 6.32 (dt, $J = 16.8, 10.0$ Hz, 1H), 5.20–5.09 (m, 2H), 4.41 (d, $J = 9.9$ Hz, 1H), 3.75 (s, 3H), 3.56 (s, 3H), 3.23 (ddd, $J = 17.1, 12.2, 4.6$ Hz, 1H), 2.88 (ddd, $J = 17.6, 4.9, 2.9$ Hz, 1H), 2.58 (ddd, $J = 13.6, 4.7, 3.0$ Hz, 1H), 2.11 (ddd, $J = 13.6, 12.2, 5.0$ Hz, 1H); ^{13}C NMR (126 MHz, $CDCl_3$) δ 193.3, 170.1, 158.4, 143.2, 136.8, 133.6, 132.5, 131.8, 131.2, 128.8, 128.4, 126.7, 117.6, 113.5, 62.8, 55.3, 53.2, 52.6, 28.7, 26.4; IR (Neat Film, NaCl) 3073, 3003, 2950, 2836, 1732, 1687, 1636, 1608, 1601, 1581, 1511, 1454, 1442, 1435, 1357, 1337, 1303, 1242, 1215, 1181, 1114, 1078, 1033, 1000, 981, 923, 893, 834, 808, 749 cm^{-1} ; HRMS (ESI+) m/z calc'd for fragment $C_{10}H_{11}O$ [$M-$

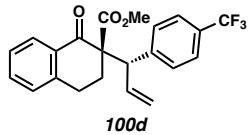
$C_{11}H_{12}O_3+H]^+$: 147.0804, found 147.0807; HPLC conditions: 2% IPA, 0.6 mL/min, Chiralpak AD-H column, λ = 254 nm, t_R (min): minor = 27.44, major = 37.29.

(R)-methyl 2-((S)-1-(4-bromophenyl)allyl)-1-oxo-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100c)



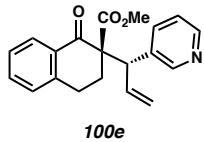
Ketoester **100c** was isolated by silica gel chromatography (gradient elution, 0→3% EtOAc in hexanes) as a colorless oil. 99% ee, $[\alpha]_D^{25} +49.1$ (c 1.18, $CHCl_3$); R_f = 0.4 (5% EtOAc in hexanes); 1H NMR (500 MHz, $CDCl_3$) δ 8.02 (dd, J = 8.0, 1.4 Hz, 1H), 7.43 (td, J = 7.5, 1.5 Hz, 1H), 7.39–7.35 (m, 2H), 7.32–7.27 (m, 2H), 7.27–7.25 (m, 1H), 7.15 (dt, J = 7.7, 0.9 Hz, 1H), 6.29 (dt, J = 16.7, 10.0 Hz, 1H), 5.32–5.03 (m, 2H), 4.37 (d, J = 9.9 Hz, 1H), 3.54 (s, 3H), 3.29–3.15 (m, 1H), 2.88 (ddd, J = 17.5, 4.9, 2.8 Hz, 1H), 2.57 (ddd, J = 13.6, 4.7, 2.9 Hz, 1H), 2.09 (ddd, J = 13.5, 12.3, 4.9 Hz, 1H); ^{13}C NMR (126 MHz, $CDCl_3$) δ 193.2, 169.9, 143.0, 139.0, 136.1, 133.8, 132.4, 132.0, 131.2, 128.8, 128.4, 126.8, 121.0, 118.5, 62.5, 53.7, 52.7, 29.1, 26.4; IR (Neat Film, NaCl) 3074, 3025, 2949, 1732, 1687, 1683, 1633, 1601, 1488, 1454, 1435, 1403, 1357, 1297, 1240, 1215, 1170, 1141, 1112, 1075, 1032, 1010, 981, 925, 892, 831, 808, 750, 741 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $C_{21}H_{20}^{79}BrO_3$ $[M+H]^+$: 399.0590, found 399.0585; HPLC conditions: 2% IPA, 0.6 mL/min, Chiralpak AD-H column, λ = 254 nm, t_R (min): minor = 19.71, major = 23.59.

(R)-methyl 1-oxo-2-((S)-1-(4-(trifluoromethyl)phenyl)allyl)-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100d)



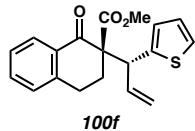
Ketoester **100d** was isolated by silica gel chromatography (gradient elution, 0→5% EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25} +32.4$ (c 1.51, CHCl_3); $R_f = 0.3$ (5% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.03 (dd, $J = 7.9, 1.4$ Hz, 1H), 7.57 (d, $J = 8.4$ Hz, 2H), 7.52 (d, $J = 8.3$ Hz, 2H), 7.44 (td, $J = 7.5, 1.4$ Hz, 1H), 7.28 (t, $J = 7.5$, 1H), 7.16 (d, $J = 7.7$ Hz, 1H), 6.34 (dt, $J = 16.7, 10.1$ Hz, 1H), 5.33–5.08 (m, 2H), 4.45 (d, $J = 10.0$ Hz, 1H), 3.54 (s, 3H), 3.29–3.16 (m, 1H), 2.90 (ddd, $J = 17.6, 4.9, 2.7$ Hz, 1H), 2.60 (ddd, $J = 13.6, 4.7, 2.8$ Hz, 1H), 2.11 (ddd, $J = 13.5, 12.3, 5.0$ Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 193.1, 169.9, 144.1, 143.0, 135.8, 133.8, 132.4, 130.6, 129.1 (q, $^2J_{\text{CF}} = 32.4$ Hz), 128.8, 128.4, 126.8, 125.0 (q, $^3J_{\text{CF}} = 3.8$ Hz), 124.3 (q, $^1J_{\text{CF}} = 272.0$ Hz), 118.8, 62.5, 54.1, 52.7, 29.3, 26.4; IR (Neat Film, NaCl) 3074, 2952, 1736, 1733, 1689, 1683, 1616, 1601, 1454, 1435, 1413, 1327, 1241, 1217, 1166, 1123, 1070, 1019, 927, 846, 809, 751, 742 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{22}\text{H}_{20}\text{F}_3\text{O}_3$ $[\text{M}+\text{H}]^+$: 389.1359, found 389.1346; SFC conditions: 5% IPA, 4.0 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): minor = 3.38, major = 3.91.

(R)-methyl 1-oxo-2-((S)-1-(pyridin-3-yl)allyl)-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100e)



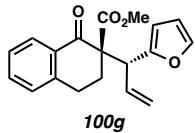
Ketoester **100e** was isolated by silica gel chromatography (gradient elution 20→50% EtOAc in hexanes) as a white solid. 98% ee, $[\alpha]_D^{25} +64.6$ (c 0.46, CHCl_3); $R_f = 0.4$ (50% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.60 (s, 1H), 8.43 (dd, $J = 5.0, 1.7$ Hz, 1H), 8.02 (dd, $J = 8.3, 1.3$ Hz, 1H), 7.93 (dt, $J = 8.0, 2.0$ Hz, 1H), 7.44 (td, $J = 7.5, 1.5$ Hz, 1H), 7.28 (t, $J = 7.6$ Hz, 1H), 7.23 (dd, $J = 8.0, 4.8$ Hz, 1H), 7.17 (d, $J = 7.7$ Hz, 1H), 6.36–6.29 (m, 1H), 5.22–5.18 (m, 2H), 4.31 (d, $J = 9.8$ Hz, 1H), 3.53 (s, 3H), 3.21 (ddd, $J = 17.2, 12.3, 4.7$ Hz, 1H), 2.91 (ddd, $J = 17.5, 4.9, 2.7$ Hz, 1H), 2.60 (ddd, $J = 13.5, 4.7, 2.8$ Hz, 1H), 2.17 (ddd, $J = 13.4, 12.3, 4.9$ Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 193.3, 170.0, 150.9, 147.9, 142.9, 138.4, 136.0, 135.5, 133.9, 132.5, 128.8, 128.4, 126.9, 123.3, 119.2, 62.4, 52.7, 52.5, 29.7, 26.5; IR (Neat Film, NaCl) 3029, 2950, 2848, 1732, 1687, 1599, 1573, 1479, 1454, 1429, 1356, 1295, 1274, 1241, 1216, 1171, 1122, 1077, 1025, 999, 979, 926, 807, 749, 716 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{20}\text{H}_{20}\text{NO}_3$ $[\text{M}+\text{H}]^+$: 322.1438, found 322.1442; HPLC conditions: 10% IPA, 1.0 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): minor = 13.45, major = 15.72.

(R)-methyl 1-oxo-2-((R)-1-(thiophen-2-yl)allyl)-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100f)



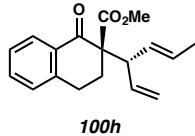
Ketoester **100f** was isolated by silica gel chromatography (gradient elution, 0→3% EtOAc in hexanes) as a white solid. 95% ee, $[\alpha]_D^{25} -14.2$ (*c* 0.86, CHCl₃); *R*_f = 0.4 (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 8.08 (dd, *J* = 7.9, 1.2 Hz, 1H), 7.44 (td, *J* = 7.5, 1.4 Hz, 1H), 7.29 (t, *J* = 7.6 Hz, 1H), 7.17 (d, *J* = 7.7 Hz, 1H), 7.14 (dd, *J* = 5.1, 1.2 Hz, 1H), 6.93 (ddd, *J* = 3.6, 1.2, 0.7 Hz, 1H), 6.88 (dd, *J* = 5.1, 3.5 Hz, 1H), 6.23 (dt, *J* = 16.8, 10.0 Hz, 1H), 5.26–5.12 (m, 2H), 4.76 (d, *J* = 10.0 Hz, 1H), 3.59 (s, 3H), 3.25 (ddd, *J* = 17.2, 12.0, 4.8 Hz, 1H), 2.89 (ddd, *J* = 17.5, 5.0, 3.1 Hz, 1H), 2.55 (ddd, *J* = 13.7, 4.8, 3.1 Hz, 1H), 2.12 (ddd, *J* = 13.6, 12.0, 5.0 Hz, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 193.2, 169.7, 143.2, 142.3, 135.9, 133.8, 132.3, 128.9, 128.4, 126.9, 126.8, 126.4, 124.9, 118.3, 62.9, 52.7, 49.5, 28.0, 26.2; IR (Neat Film, NaCl) 3071, 2949, 2925, 2853, 1731, 1686, 1639, 1599, 1484, 1453, 1433, 1354, 1293, 1272, 1240, 1214, 1170, 1119, 1078, 1032, 979, 924, 891, 853, 807, 749 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₁₉H₁₉SO₃ [M+H]⁺: 327.1049, found 327.1048; SFC conditions: 10% IPA, 4.0 mL/min, Chiralcel OJ-H column, λ = 254 nm, *t*_R (min): major = 2.96, minor = 3.63.

(R)-methyl 2-((R)-1-(furan-2-yl)allyl)-1-oxo-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100g)



Ketoester **100g** was isolated by silica gel chromatography (gradient elution, 0→3% EtOAc in hexanes) as a colorless oil. 95% ee, $[\alpha]_D^{25} +22.5$ (c 1.17, CHCl_3); $R_f = 0.4$ (5% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.06 (dd, $J = 8.0, 1.4$ Hz, 1H), 7.45 (td, $J = 7.5, 1.5$ Hz, 1H), 7.28 (t, $J = 7.6$ Hz, 1H), 7.25 (dd, $J = 1.9, 0.9$ Hz, 1H), 7.17 (d, $J = 7.7$ Hz, 1H), 6.25 (dd, $J = 3.2, 1.8$ Hz, 1H), 6.19–6.09 (m, 2H), 5.26–5.18 (m, 2H), 4.63 (d, $J = 9.8$ Hz, 1H), 3.63 (s, 3H), 3.24 (ddd, $J = 17.3, 12.2, 4.8$ Hz, 1H), 2.88 (ddd, $J = 17.5, 4.9, 3.1$ Hz, 1H), 2.57 (ddd, $J = 13.8, 4.8, 3.1$ Hz, 1H), 1.99 (ddd, $J = 13.8, 12.1, 5.0$ Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 192.7, 169.7, 153.4, 143.3, 141.6, 133.7, 133.6, 132.1, 128.9, 128.4, 126.7, 119.2, 110.3, 108.4, 62.2, 52.7, 47.9, 28.1, 26.1; IR (Neat Film, NaCl) 3116, 3075, 3024, 2950, 2848, 1734, 1731, 1689, 1639, 1600, 1500, 1485, 1453, 1433, 1356, 1293, 1271, 1243, 1216, 1172, 1155, 1120, 1110, 1078, 1012, 981, 965, 928, 905, 892, 806, 745, 736 cm^{-1} ; HRMS (ESI+) m/z calc'd for $\text{C}_{19}\text{H}_{19}\text{O}_4$ $[\text{M}+\text{H}]^+$: 311.1278, found 311.1275; SFC conditions: 10% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): major = 5.21, minor = 6.03.

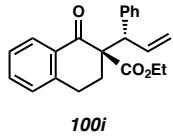
(R)-methyl 2-((S,E)-hexa-1,4-dien-3-yl)-1-oxo-1,2,3,4-tetrahydronaphthalene-2-carboxylate (100h)



Ketoester **100h** was isolated by silica gel chromatography (gradient elution, 0→2% EtOAc in hexanes) as a colorless oil. 90% ee, $[\alpha]_D^{25} +46.4$ (c 1.02, CHCl_3); $R_f = 0.5$ (5% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.05 (dd, $J = 7.9, 1.2$ Hz, 1H), 7.45 (td, $J = 7.5, 1.5$ Hz, 1H), 7.30 (t, $J = 7.6$ Hz, 1H), 7.19 (d, $J = 7.7$ Hz, 1H), 5.98–5.87 (m, 1H), 5.74 (ddd, $J = 15.3, 8.0, 1.6$ Hz, 1H), 5.57–5.48 (m, 1H), 5.08–5.03 (m, 2H), 3.61 (s, 3H), 3.47 (t, $J = 8.4$ Hz, 1H), 3.12 (ddd, $J = 17.0, 12.0, 4.7$ Hz, 1H), 2.91 (dt, $J = 17.4, 4.1$ Hz, 1H), 2.45 (ddd, $J = 13.7, 4.7, 3.3$ Hz, 1H), 2.25 (ddd, $J = 13.7, 11.9, 4.9$ Hz, 1H), 1.65 (ddd, $J = 6.4, 1.7, 0.7$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 194.2, 171.0, 143.1, 136.7, 133.6, 132.8, 129.8, 128.8, 128.2, 128.2, 126.8, 117.6, 62.0, 53.2, 52.4, 29.5, 26.5, 18.2; IR (Neat Film, NaCl) 3075, 3028, 2951, 2854, 1732, 1688, 1600, 1454, 1438, 1356, 1300, 1272, 1235, 1214, 1169, 1122, 1090, 999, 974, 917, 890, 803, 747 cm^{-1} ; HRMS (ESI+) m/z calc'd for $\text{C}_{18}\text{H}_{21}\text{O}_3$ $[\text{M}+\text{H}]^+$: 288.1485, found 288.1489; SFC conditions: 2% MeOH, 2.5 mL/min, Chiraldak IC column, $\lambda = 254$ nm, t_R (min): minor = 8.23, major = 8.87.

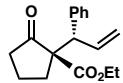
(R)-ethyl 1-oxo-2-((S)-1-phenylallyl)-1,2,3,4-tetrahydronaphthalene-2-carboxylate

(100i)



Ketoester **100i** was isolated by silica gel chromatography (gradient elution, 0→5% EtOAc in hexanes) as a white solid, >99% ee, $[\alpha]_D^{25} +42.7$ (c 1.09, CHCl_3); $R_f = 0.3$ (5% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 8.03 (dd, $J = 7.9, 1.4$ Hz, 1H), 7.46–7.39 (m, 3H), 7.29–7.23 (m, 3H), 7.20–7.12 (m, 2H), 6.37 (dt, $J = 16.8, 10.0$ Hz, 1H), 5.20–5.07 (m, 2H), 4.40 (d, $J = 9.9$ Hz, 1H), 4.07–3.94 (m, 2H), 3.22 (ddd, $J = 17.3, 12.2, 4.8$ Hz, 1H), 2.88 (ddd, $J = 17.5, 5.0, 2.9$ Hz, 1H), 2.58 (ddd, $J = 13.6, 4.7, 3.0$ Hz, 1H), 2.12 (ddd, $J = 13.6, 12.1, 5.0$ Hz, 1H), 1.06 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 193.4, 169.7, 143.0, 140.0, 136.7, 133.5, 132.7, 130.3, 128.7, 128.3, 128.1, 126.9, 126.7, 117.9, 62.3, 61.6, 54.1, 29.2, 26.4, 14.0; IR (Neat Film, NaCl) 3063, 3027, 2978, 2934, 1727, 1699, 1689, 1685, 1599, 1490, 1452, 1363, 1298, 1282, 1235, 1212, 1157, 1107, 1080, 1018, 926, 899, 787, 773, 743 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{22}\text{H}_{23}\text{O}_3$ $[\text{M}+\text{H}]^+$: 335.1642, found 335.1651; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 254$ nm, t_R (min): minor = 13.82, major = 16.53.

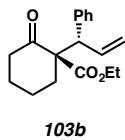
(R)-ethyl 2-oxo-1-((S)-1-phenylallyl)cyclopentanecarboxylate (103a)



103a

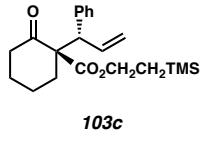
Ketoester **103a** was isolated by silica gel chromatography (gradient elution, 0→5% EtOAc in hexanes) as a colorless oil. 99% ee, $[\alpha]_D^{25} -52.5$ (c 1.04, CHCl_3); $R_f = 0.3$ (5% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.31–7.15 (m, 5H), 6.14–6.03 (m, 1H), 5.20–5.10 (m, 2H), 4.37 (d, $J = 8.9$ Hz, 1H), 4.22–4.09 (m, 2H), 2.67 (dd, $J = 13.4, 7.1, 3.5, 1.7$ Hz, 1H), 2.24–2.14 (m, 1H), 2.13–2.02 (m, 1H), 1.84–1.71 (m, 1H), 1.69–1.59 (m, 1H), 1.59–1.49 (m, 1H), 1.24 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 213.4, 169.2, 139.3, 136.3, 129.9, 128.4, 127.1, 117.8, 65.9, 61.9, 52.8, 38.9, 28.5, 19.7, 14.2; IR (Neat Film, NaCl) 3083, 3062, 3030, 2979, 2891, 1752, 1719, 1639, 1601, 1493, 1465, 1452, 1405, 1365, 1315, 1223, 1138, 1105, 1026, 1003, 923, 864, 826, 757, 707 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{17}\text{H}_{21}\text{O}_3$ $[\text{M}+\text{H}]^+$: 273.1485, found 273.1483; HPLC conditions: 2% IPA, 0.6 mL/min, Chiralcel OD-H column, $\lambda = 210$ nm, t_R (min): major = 11.23, minor = 12.73.

(R)-ethyl 2-oxo-1-((S)-1-phenylallyl)cyclohexanecarboxylate (103b)



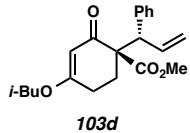
Ketoester **103b** was isolated by silica gel chromatography (gradient elution, 0→5% EtOAc in hexanes) as a white solid. 98% ee, $[\alpha]_D^{25} +140.6$ (*c* 1.25, CHCl_3); $R_f = 0.4$ (5% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.36–7.30 (m, 2H), 7.29–7.22 (m, 2H), 7.21–7.15 (m, 1H), 6.33 (ddd, *J* = 16.9, 10.2, 9.2 Hz, 1H), 5.13–4.99 (m, 2H), 4.11–3.96 (m, 2H), 3.93 (d, *J* = 9.2 Hz, 1H), 2.47–2.39 (m, 2H), 2.36–2.29 (m, 1H), 1.92 (dddd, *J* = 9.5, 4.8, 2.7, 1.5 Hz, 1H), 1.79–1.47 (m, 3H), 1.12 (t, *J* = 7.1 Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 206.7, 170.8, 140.0, 137.5, 130.3, 128.0, 126.8, 117.4, 65.5, 61.4, 54.6, 42.1, 35.0, 27.2, 22.8, 14.0; IR (Neat Film, NaCl) 3077, 3028, 2977, 2939, 2865, 1714, 1635, 1600, 1491, 1452, 1388, 1365, 1340, 1309, 1262, 1231, 1204, 1133, 1085, 1020, 1002, 919, 854, 756, 704 cm^{-1} ; HRMS (MM: ESI-APCI+) *m/z* calc'd for $\text{C}_{18}\text{H}_{23}\text{O}_3$ $[\text{M}+\text{H}]^+$: 287.1642, found 287.1639; SFC conditions: 10% IPA, 4.0 mL/min, Chiralpak IC column, $\lambda = 210$ nm, t_R (min): minor = 1.69, major = 1.94.

(R)-2-(trimethylsilyl)ethyl 2-oxo-1-((S)-1-phenylallyl)cyclohexanecarboxylate (103c)



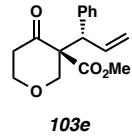
Ketoester **103c** was isolated by silica gel chromatography (gradient elution, 0→2% i-BuOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25} +91.6$ (c 0.45, CHCl_3); $R_f = 0.4$ (10% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.37–7.32 (m, 2H), 7.24 (ddd, J = 8.2, 6.9, 1.4 Hz, 2H), 7.21–7.15 (m, 1H), 6.34 (ddd, J = 16.9, 10.2, 9.2 Hz, 1H), 5.21–4.86 (m, 2H), 4.05 (dddd, J = 58.0, 11.8, 10.8, 5.9 Hz, 2H), 3.92 (d, J = 9.2 Hz, 1H), 2.46–2.40 (m, 2H), 2.38–2.28 (m, 1H), 1.98–1.87 (m, 1H), 1.80–1.71 (m, 1H), 1.72–1.47 (m, 3H), 0.82 (qdd, J = 13.6, 11.7, 5.7 Hz, 2H), 0.01 (s, 9H); ^{13}C NMR (126 MHz, CDCl_3) δ 206.6, 170.9, 139.9, 137.4, 130.2, 127.9, 126.7, 117.3, 65.4, 63.7, 54.5, 42.0, 34.9, 27.0, 22.7, 17.2, –1.6; IR (Neat Film, NaCl) 2950, 1712, 1452, 1250, 1231, 1133, 921, 859, 837 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{21}\text{H}_{30}\text{NaO}_3\text{Si}$ $[\text{M}+\text{Na}]^+$: 381.1856, found 381.1865; SFC conditions: 3% IPA, 2.5 mL/min, Chiralcel OJ-H column, $\lambda = 210$ nm, t_R (min): minor = 1.93, major = 2.24.

(R)-methyl 4-isobutoxy-2-oxo-1-((S)-1-phenylallyl)cyclohex-3-enecarboxylate (103d)



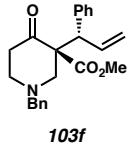
Ketoester **103d** was isolated by silica gel chromatography (gradient elution, 0→10% EtOAc in hexanes) as a colorless oil. >99% ee, $[\alpha]_D^{25} +31.5$ (c 1.88, CHCl_3); $R_f = 0.4$ (10% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.34–7.29 (m, 2H), 7.27–7.20 (m, 2H), 7.20–7.13 (m, 1H), 6.24 (dt, $J = 16.7, 10.2$ Hz, 1H), 5.28 (s, 1H), 5.18 (ddd, $J = 16.8, 1.9, 0.8$ Hz, 1H), 5.12 (dd, $J = 10.0, 1.8$ Hz, 1H), 4.62 (d, $J = 10.4$ Hz, 1H), 3.64 (s, 3H), 3.53–3.45 (m, 2H), 2.79 (dddd, $J = 18.3, 11.8, 5.1, 1.6$ Hz, 1H), 2.43 (ddd, $J = 13.4, 5.1, 2.4$ Hz, 1H), 2.26 (ddd, $J = 18.2, 5.5, 2.4$ Hz, 1H), 1.94 (dt, $J = 13.3, 6.7$ Hz, 1H), 1.83 (ddd, $J = 13.4, 11.8, 5.5$ Hz, 1H), 0.91 (d, $J = 6.8$ Hz, 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 192.8, 177.2, 170.0, 139.7, 136.1, 130.2, 128.1, 126.8, 118.0, 102.9, 74.9, 61.1, 52.6, 52.1, 27.7, 26.7, 25.2, 19.11, 19.08; IR (Neat Film, NaCl) 3073, 3029, 2958, 2874, 1727, 1664, 1607, 1582, 1491, 1470, 1452, 1443, 1431, 1406, 1384, 1369, 1316, 1298, 1231, 1193, 1177, 1140, 1116, 1079, 1012, 987, 921, 903, 844, 817, 788, 764, 724 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{21}\text{H}_{27}\text{O}_4$ $[\text{M}+\text{H}]^+$: 343.1904, found 343.1905; SFC conditions: 10% IPA, 2.5 mL/min, Chiralcel OD-H column, $\lambda = 254$ nm, t_R (min): major = 3.71, minor = 6.24.

(S)-methyl 4-oxo-3-((S)-1-phenylallyl)tetrahydro-2H-pyran-3-carboxylate (103e)



Ketoester **103e** was isolated by silica gel chromatography (gradient elution, 5→10% EtOAc in hexanes) as a colorless oil. 98% ee, $[\alpha]_D^{25} +71.1$ (c 0.88, CHCl_3); $R_f = 0.2$ (10% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.35–7.30 (m, 2H), 7.31–7.25 (m, 2H), 7.24–7.19 (m, 1H), 6.43 (ddd, $J = 16.9, 10.2, 9.4$ Hz, 1H), 5.19–5.03 (m, 2H), 4.28 (dd, $J = 11.9, 1.2$ Hz, 1H), 4.03 (dddd, $J = 11.1, 6.2, 4.9, 1.3$ Hz, 1H), 3.98 (d, $J = 9.4$ Hz, 1H), 3.82 (dddd, $J = 11.3, 9.0, 4.5, 0.6$ Hz, 1H), 3.67 (d, $J = 11.8$ Hz, 1H), 3.61 (s, 3H), 2.70 (ddd, $J = 14.5, 8.9, 6.2$ Hz, 1H), 2.57 (dt, $J = 14.5, 4.7$ Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 202.8, 169.7, 138.6, 136.7, 129.8, 128.4, 127.4, 118.0, 73.2, 68.6, 67.1, 52.4, 51.6, 41.9; IR (Neat Film, NaCl) 3063, 3029, 2973, 2951, 2863, 1746, 1716, 1635, 1600, 1492, 1472, 1454, 1433, 1378, 1360, 1310, 1290, 1229, 1212, 1176, 1140, 1112, 1085, 1033, 1001, 978, 925, 826, 763, 741 cm^{-1} ; HRMS (ESI+) m/z calc'd for $\text{C}_{16}\text{H}_{19}\text{O}_4$ $[\text{M}+\text{H}]^+$: 275.1278, found 275.1282; SFC conditions: 5% IPA, 2.5 mL/min, Chiralpak AD-H column, $\lambda = 210$ nm, t_R (min): minor = 4.65, major = 4.95.

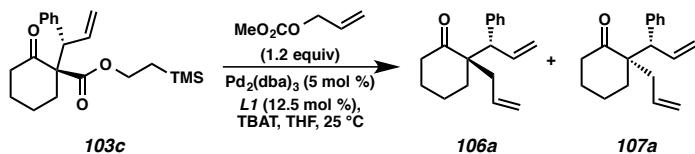
(S)-methyl 1-benzyl-4-oxo-3-((S)-1-phenylallyl)piperidine-3-carboxylate (103f)



103f

Ketoester **103f** was isolated by silica gel chromatography (gradient elution, 5→10% EtOAc in hexanes) as a colorless oil. 97% ee, $[\alpha]_D^{25} +34.3$ (c 0.87, CHCl_3); $R_f = 0.3$ (10% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.39–7.29 (m, 5H), 7.28–7.24 (m, 2H), 7.24–7.14 (m, 3H), 6.44 (ddd, $J = 16.9, 10.2, 9.3$ Hz, 1H), 5.12–5.04 (m, 2H), 4.03 (d, $J = 9.4$ Hz, 1H), 3.59 (s, 3H), 3.64–3.54 (m, 2H), 3.15 (dd, $J = 11.9, 2.0$ Hz, 1H), 2.82–2.75 (m, 1H), 2.71 (ddd, $J = 14.1, 8.5, 5.7$ Hz, 1H), 2.65–2.56 (m, 2H), 2.53 (ddd, $J = 13.9, 5.4, 4.3$ Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 205.3, 170.6, 139.4, 138.0, 137.4, 129.9, 129.2, 128.5, 128.2, 127.5, 127.1, 117.6, 66.1, 62.0, 60.2, 53.4, 53.0, 52.0, 40.9; IR (Neat Film, NaCl) 3060, 3027, 2949, 2811, 2765, 1718, 1631, 1600, 1584, 1493, 1468, 1452, 1432, 1364, 1345, 1310, 1286, 1228, 1194, 1138, 1073, 1047, 1028, 1001, 973, 922, 821, 740 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{23}\text{H}_{26}\text{NO}_3$ $[\text{M}+\text{H}]^+$: 364.1907, found 364.1908; SFC conditions: 10% IPA, 4.0 mL/min, Chiralpak AD-H column, $\lambda = 210$ nm, t_R (min): major = 2.49, minor = 2.94.

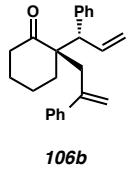
3.6.5. General procedure for Pd-catalyzed allylic alkylation



(R)-2-allyl-2-((S)-1-phenylallyl)cyclohexanone (106a). To a 0.5 dram scintillation vial equipped with a magnetic stir bar were added $\text{Pd}_2(\text{dba})_3$ (1.3 mg, 0.0014 mmol), **L8** (1.2 mg, 0.0035 mmol), TBAT (16.6 mg, 0.031 mmol) and THF (0.9 mL) in a nitrogen-filled glove box. The dark purple mixture was stirred at ambient glove box temperature (ca. 30 °C) for 35 minutes at which point the mixture had become red-orange. Ketoester **103c** (10.0 mg, 0.028 mmol) and allyl methylcarbonate (4.1 mg, 0.035 mmol) were then added neat to the reaction mixture. The resulting yellow-green reaction mixture was stirred at 20 °C until full conversion of the starting material was indicated by TLC analysis (reaction times typically ranged from 24 to 36 hours). The vial was removed from the glove box, uncapped and diluted with 2 ml of hexanes. Filtration through a celite pad afforded the crude residue, which was concentrated *in vacuo* and analyzed by ^1H NMR to determine the diastereomeric ratio of **106a** and **107a** (2:1). The residue was purified by silica gel flash chromatography (gradient elution, 0→2% EtOAc in hexanes) to afford **106a** and **107a** (6.5 mg, 91% combined yield) as a colorless oil. 99% ee (The enantiomeric excesses of the products **106a** and **107a** are inferred from the corresponding Ir-catalyzed allylic alkylation products (**103c**)). Spectroscopic data for compound **106a** is as follows: $[\alpha]_D^{25} -1.9$ (*c* 0.48, CHCl_3); $R_f = 0.3$ (0.4% EtOAc in hexanes); ^1H NMR (300 MHz, CDCl_3) δ 7.33–7.28 (m, 2H), 7.25–7.20 (m, 3H), 6.20 (dt, $J = 16.8, 10.1$ Hz, 1H),

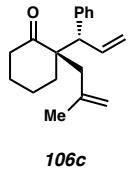
5.68 (dddd, $J = 17.1, 10.1, 8.8, 5.4$ Hz, 1H), 5.09 (ddd, $J = 10.1, 1.6, 0.4$ Hz, 1H), 5.08 (ddd, $J = 16.7, 1.7, 0.8$ Hz, 1H), 4.98 (dddd, $J = 10.2, 2.3, 1.3, 0.7$ Hz, 1H), 4.92–4.86 (m, 1H), 3.92 (d, $J = 9.9$ Hz, 1H), 2.75 (dd, $J = 13.7, 5.4$ Hz, 1H), 2.56–2.10 (m, 2H), 2.11–1.59 (m, 7H); ^{13}C NMR (126 MHz, CDCl_3) δ 212.9, 140.1, 136.3, 134.9, 130.0, 128.1, 126.8, 117.9, 117.4, 55.7, 52.8, 40.5, 37.6, 31.9, 26.2, 21.1; IR (Neat Film, NaCl) 3073, 3028, 2937, 2864, 1833, 1701, 1636, 1600, 1452, 1432, 1313, 1219, 1125, 1056, 1002, 916, 849, 787, 765 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{18}\text{H}_{23}\text{O}$ [M+H] $^+$: 255.1754; found 255.1743. Spectroscopic data for compound **107a** is as follows: $[\alpha]_D^{25}$ +6.1 (c 0.75, CHCl_3); $R_f = 0.3$ (0.4% EtOAc in hexanes); ^1H NMR (300 MHz, CDCl_3) δ 7.33–7.28 (m, 2H), 7.25–7.20 (m, 3H), 6.19 (dt, $J = 16.8, 10.1$ Hz, 1H), 5.65 (dddd, $J = 17.1, 10.1, 8.8, 5.4$ Hz, 1H), 5.17 (dd, $J = 1.7, 1.0$, 1H), 5.15 (dd, $J = 1.7, 1.0$, 1H) 5.07–5.02 (m, 2H) 3.95 (d, $J = 8.5$, 1H), 2.75 (dd, $J = 13.7, 5.4$ Hz, 1H), 2.56–2.10 (m, 2H), 2.11–1.59 (m, 7H); ^{13}C NMR (126 MHz, CDCl_3) δ 213.2, 140.3, 138.0, 134.1, 129.8, 128.3, 126.8, 118.0, 117.8, 55.6, 53.4, 40.6, 38.6, 32.2, 25.6, 20.8; IR (Neat Film, NaCl) 3073, 3028, 2937, 2864, 1833, 1701, 1636, 1600, 1452, 1432, 1313, 1219, 1125, 1056, 1002, 916, 849, 787, 765 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{18}\text{H}_{23}\text{O}$ [M+H] $^+$: 255.1754; found 255.1750.

(R)-2-((S)-1-phenylallyl)-2-(2-phenylallyl)cyclohexanone (106b)



Ketone **106b** was isolated by silica gel chromatography (gradient elution, 0→1% Et₂O in hexanes) as a colorless oil. $[\alpha]_D^{25} -50.9$ (*c* 0.22, CHCl₃); $R_f = 0.5$ (10% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.32–7.13 (m, 10H), 6.24 (dt, *J* = 16.8, 10.1 Hz, 1H), 5.29 (d, *J* = 1.8 Hz, 1H), 5.22–5.02 (m, 3H), 3.92 (d, *J* = 10.0 Hz, 1H), 2.95 (ddd, *J* = 377.8, 13.8, 0.9 Hz, 2H), 2.15–2.00 (m, 2H), 1.76–1.57 (m, 4H), 1.54–1.43 (m, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 212.8, 145.6, 142.8, 140.3, 136.2, 129.9, 128.1, 128.0, 127.0, 126.7, 126.5, 118.2, 117.7, 56.6, 54.4, 40.6, 37.6, 30.6, 24.8, 21.4; IR (Neat Film, NaCl) 2937, 2859, 1701, 1624, 1597, 1451, 1310, 1256, 1207, 1125, 910, 779 cm⁻¹; HRMS (MM: ESI-APCI+) *m/z* calc'd for C₂₄H₂₆O [M+H]⁺: 331.2056, found 331.2065.

(R)-2-(2-methylallyl)-2-((S)-1-phenylallyl)cyclohexanone (106c)

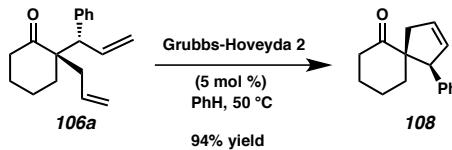


Ketone **106c** was isolated by silica gel chromatography (gradient elution, 0→2% EtOAc in hexanes) as a colorless oil. $[\alpha]_D^{25} -41.7$ (*c* 0.46, CHCl₃); $R_f = 0.4$ (5% EtOAc in hexanes); ¹H NMR (500 MHz, CDCl₃) δ 7.33–7.28 (m, 2H), 7.25–7.19 (m, 3H), 6.19 (dt,

$J = 16.8, 10.1$ Hz, 1H), 5.08 (dd, $J = 10.2, 1.6$ Hz, 1H), 5.03 (ddd, $J = 16.8, 1.6, 0.8$ Hz, 1H), 4.74 (ddt, $J = 2.7, 1.8, 0.9$ Hz, 1H), 4.52 (ddt, $J = 2.4, 1.6, 0.9$ Hz, 1H), 3.86 (d, $J = 10.0$ Hz, 1H), 2.90 (d, $J = 13.1$ Hz, 1H), 2.37 (dtd, $J = 16.2, 4.9, 1.6$ Hz, 1H), 2.25 (ddd, $J = 15.9, 11.3, 6.0$ Hz, 1H), 2.00 (d, $J = 13.7$ Hz, 1H), 1.93–1.78 (m, 2H), 1.74–1.63 (m, 4H), 1.59 (dt, $J = 1.4, 0.7$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 212.8, 143.0, 140.3, 136.5, 130.1, 128.2, 126.9, 117.4, 115.3, 55.5, 54.2, 40.7, 40.2, 30.8, 25.5, 25.1, 21.4; IR (Neat Film, NaCl) 3071, 3030, 2940, 2865, 1704, 1637, 1599, 1452, 1375, 1314, 1209, 1124, 994, 916, 893, 756 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{19}\text{H}_{25}\text{O}$ [M+H] $^+$: 269.1920; found 269.1920.

3.6.6. Determination of the relative configuration of compound 106a

The relative configuration of compound **106a** was determined via NOE analysis of the corresponding spirocycle, **108**, obtained via ring-closing metathesis. The experimental procedure by which **108** was generated is as follows:



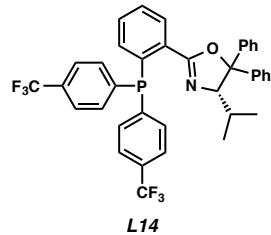
(1*S*,5*R*)-1-phenylspiro[4.5]dec-2-en-6-one (108). To a flask charged with Grubbs-Hoveyda second generation catalyst (1.85 mg, 0.0030 mmol) under an atmosphere of argon was added a solution of cyclohexanone **106a** (15.0 mg, 0.059 mmol) in 6 mL benzene. The reaction mixture was heated to 50 °C and stirred for 4 hours, at which point the reaction was determined to be complete by TLC analysis. The reaction vessel was

cooled to 25 °C and 0.5 mL of ethyl vinyl ether was added. After 30 minutes of stirring, the crude mixture was purified directly by silica gel chromatography (gradient elution, 0→3% EtOAc in hexanes) to afford spirocycle **108** (12.7 mg, 0.056 mmol, 94% yield) as a colorless oil. $[\alpha]_D^{25} -133.6$ (*c* 0.25, CHCl_3); $R_f = 0.5$ (10% EtOAc in hexanes); ^1H NMR (500 MHz, CDCl_3) δ 7.32–7.24 (m, 2H), 7.24–7.18 (m, 1H), 7.18–7.12 (m, 2H), 6.01–5.52 (m, 1H), 4.75 (p, *J* = 2.1 Hz, 1H), 2.66–2.56 (m, 2H), 2.56–2.44 (m, 2H), 1.95–1.85 (m, 1H), 1.64–1.49 (m, 3H), 1.42 (dtd, *J* = 14.66, 3.6, 2.3 Hz, 1H), 1.35–0.72 (m, 1H), 1.01 (ddd, *J* = 13.9, 11.3, 4.5 Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 212.9, 140.6, 133.8, 129.3, 128.0, 127.4, 126.4, 59.8, 53.7, 42.8, 39.6, 35.5, 26.8, 22.2; IR (Neat Film, NaCl) 3944, 3693, 3053, 2986, 2941, 2866, 2685, 2305, 1698, 1422, 1264, 1129, 896, 756 cm^{-1} ; HRMS (MM: ESI-APCI+) *m/z* calc'd for $\text{C}_{16}\text{H}_{19}\text{O}$ [M+H]⁺: 227.1430; found 227.1431.

3.6.7. Spectroscopic data for new phosphinoxazoline ligands

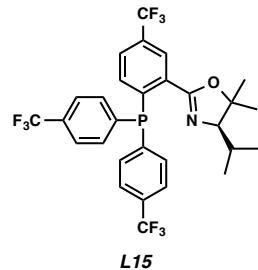
Phosphinoxazoline ligands **L14** and **L15** employed in these studies were generously donated by Dr. Robert A. Craig. They were generated following the method reported by McDougal and Stoltz. Comprehensive preparatory detail can be found in the Thesis of Dr. Craig.

(S)-2-(2-(bis(4-(trifluoromethyl)phenyl)phosphino)phenyl)-4-isopropyl-5,5-diphenyl-4,5-dihydrooxazole (L14)



$[\alpha]_D^{25} -163.33$ (c 0.75, CHCl_3); $R_f = 0.3$ (4:1 hexanes in dichloromethane); ^1H NMR (500 MHz, C_6D_6) δ 8.22 (ddd, $J = 7.8, 3.9, 1.4$ Hz, 1H), 7.29 (ddd, $J = 49.4, 8.4, 1.4$ Hz, 4H), 7.21–7.14 (m, 5H), 7.10–7.02 (m, 7H), 7.02–6.94 (m, 3H), 6.89 (td, $J = 7.6, 1.4$ Hz, 1H), 6.81 (ddd, $J = 7.9, 3.4, 1.3$ Hz, 1H), 4.66 (d, $J = 4.5$ Hz, 1H), 1.74 (td, $J = 6.6, 4.6$ Hz, 1H), 0.83 (d, $J = 6.7$ Hz, 3H), 0.59 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (126 MHz, C_6D_6) δ 160.1 (d, $J_{\text{CP}} = 3.2$ Hz), 145.8, 144.4–143.9 (m) 141.2, 137.9 (d, $J_{\text{CP}} = 27.4$ Hz), 134.9, 134.5 (dd, $J_{\text{CP}} = 70.9, 20.8$ Hz), 132.6 (d, $J_{\text{CP}} = 21.4$ Hz), 131.3, 130.6 (dd, $J_{\text{CP}} = 32.3, 18.4$ Hz), 130.3 (d, $J_{\text{CP}} = 3.2$ Hz), 129.1, 127.5 (d, $J_{\text{CF}} = 11.2$ Hz), 126.8, 125.3 (ddt, $J_{\text{CF}} = 14.7, 7.6, 3.8$ Hz), 124.8 (d, $J_{\text{CF}} = 273.5$ Hz), 93.1, 81.1 (d $J_{\text{CP}} = 2.0$ Hz), 30.60, 22.0; ^{19}F NMR (282 MHz, C_6D_6) δ -62.44, -62.53; ^{31}P NMR (121 MHz, C_6D_6) δ -7.59; IR (Neat Film, NaCl) 3060, 2961, 1654, 1605, 1493, 1470, 1448, 1396, 1323, 1166, 1127, 1060, 1016, 954, 832, 756 cm^{-1} ; HRMS (MM: ESI-APCI+) m/z calc'd for $\text{C}_{28}\text{H}_{30}{^{19}\text{F}_6}\text{NOP} [\text{M}+\text{H}]^+$: 662.2042, found 662.2080.

(R)-2-(2-(bis(4-(trifluoromethyl)phenyl)phosphino)-5-(trifluoromethyl)phenyl)-4-isopropyl-5,5-dimethyl-4,5-dihydrooxazole (L15)



$[\alpha]_D^{25} +9.45$ (*c* 3.20, CHCl_3); $R_f = 0.3$ (4:1 hexanes in dichloromethane); ^1H NMR (500 MHz, C_6D_6) δ 8.57 (dd, $J = 3.3, 2.0$ Hz, 1H), 7.41–7.36 (m, 4H), 7.21–7.15 (m, 4H), 7.10 (dd, $J = 8.2, 2.0$ Hz, 1H), 6.78 (dd, $J = 8.0, 3.0$ Hz, 1H), 3.22 (d, $J = 8.4$ Hz, 1H), 1.55 (ddt, $J = 13.0, 8.3, 6.5$ Hz, 1H), 1.21 (s, 3H), 1.08 (s, 3H), 0.99 (d, $J = 6.5$ Hz, 3H), 0.75 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (126 MHz, C_6D_6) δ 159.1 (d, $J_{\text{CP}} = 4.0$ Hz), 143.5 (t, $J_{\text{CP}} = 14.8$ Hz), 142.7 (d, $J_{\text{CP}} = 30.6$ Hz), 134.5 (dd, $J_{\text{CP}} = 21.3, 15.7$ Hz), 133.7 (d, $J_{\text{CP}} = 19.5$ Hz), 131.1 (q, $J_{\text{CF}} = 3.6$ Hz), 126.4–126.1 (m), 125.9 (d, $J_{\text{CF}} = 3.2$ Hz), 126.5–125.0 (m), 123.8 (d, $J_{\text{CF}} = 3.3$ Hz), 123.3, 87.2, 81.7 (d, $J_{\text{CP}} = 1.5$ Hz), 29.1, 28.8, 21.1, 20.8, 20.8 (d, $J_{\text{CP}} = 1.8$ Hz); ^{19}F NMR (282 MHz, C_6D_6) δ -62.63, -62.85; ^{31}P NMR (121 MHz, C_6D_6) δ -7.10; IR (Neat Film, NaCl) 2974, 1652, 1397, 1323, 1165, 1128, 1060, 1017, 832, 756 cm^{-1} ; HRMS (FAB+) *m/z* calc'd for $\text{C}_{29}\text{H}_{26}\text{O}^{19}\text{F}_9\text{NP} [\text{M}+\text{H}]^+$: 606.1608, found 606.1585.

3.6.8 Determination of enantiomeric excess

Table 3.6.8.1. Determination of enantiomeric excess

entry	compound	analytic conditions	ee (%)
1		HPLC Chiralcel OD-H, $\epsilon \approx 254$ nm 2% IPA/hexanes, 0.6 mL/min t_R (min): major 13.80, minor 17.89	>99
2		HPLC Chiralpak AD-H, $\epsilon \approx 254$ nm 2% IPA/hexanes, 0.6 mL/min t_R (min): minor 27.44, major 37.29	>99
3		HPLC Chiralpak AD-H, $\epsilon \approx 254$ nm 2% IPA/hexanes, 0.6 mL/min t_R (min): minor 19.71, major 23.59	99
4		SFC Chiralpak AD-H, $\epsilon \approx 254$ nm 5% IPA/CO ₂ , 4.0 mL/min t_R (min): minor 3.38, major 3.91	>99
5		HPLC Chiralpak AD-H, $\epsilon \approx 254$ nm 90% IPA/hexanes, 1.0 mL/min t_R (min): minor 13.45, major 15.72	98
6		SFC Chiralcel OJ-H, $\epsilon \approx 254$ nm 10% IPA/CO ₂ , 4.0 mL/min, t_R (min): major 2.96, minor 3.63	95
7		SFC Chiralpak AD-H, $\epsilon \approx 254$ nm 10% IPA/CO ₂ , 2.5 mL/min, t_R (min): major 5.21, minor 6.03	95
8		SFC Chiralpak IC, $\epsilon \approx 254$ nm 2% MeOH/CO ₂ , 2.5 mL/min, t_R (min): minor 8.23, major 8.87	90

Table 3.6.8.2 Determination of enantiomeric excess continued

entry	compound	analytic conditions	ee (%)
9		SFC Chiralpak AD-H, $\epsilon \approx 254$ nm 5% IPA/CO ₂ , 2.5 mL/min, t_R (min): minor 13.82, major 16.53	>99
10		HPLC Chiralcel OD-H, $\epsilon \approx 220$ nm 2% IPA/hexanes, 0.6 mL/min t_R (min): major 11.23, minor 12.73	99
11		SFC Chiralpak IC, $\epsilon \approx 210$ nm 10% IPA/CO ₂ , 4.0 mL/min t_R (min): minor 1.69, major 1.94	98
12		SFC Chiralcel OJ-H, $\epsilon \approx 210$ nm 3% IPA/CO ₂ , 2.5 mL/min t_R (min): minor 1.93, major 2.24	>99
13		SFC Chiralcel OD-H, $\epsilon \approx 254$ nm 10% IPA/CO ₂ , 2.5 mL/min t_R (min): major 3.71, minor 6.24	>99
14		SFC Chiralpak AD-H, $\epsilon \approx 210$ nm 5% IPA/CO ₂ , 2.5 mL/min t_R (min): minor 4.65, major 4.95	98
15		SFC Chiralpak AD-H, $\epsilon \approx 210$ nm 10% IPA/CO ₂ , 4.0 mL/min t_R (min): major 2.49, minor 2.94	97

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